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<http://www.pediatricurologycasereports.com>**Irregular bladder smooth muscle actin-gamma2 expression in *ACTG2* mutation-associated Megacystis Microcolon Intestinal Hypoperistalsis Syndrome (MMIHS): A case report****Peter Heinz-Erian^{1*}, Elisabeth Bruder², Andreas Janecke¹, Peter Rehder³, Heinz Zoller⁴, Bettina Zelger⁵, Markus Pirklbauer⁶, Thomas Müller**¹Department of Pediatric and Adolescent Medicine, Medical University of Innsbruck, Innsbruck Austria²Department of Pathology, University Hospital Basel, Basel, Switzerland³Department of Urology, Medical University of Innsbruck, Innsbruck, Austria⁴Department of Medicine-I, Medical University of Innsbruck, Innsbruck, Austria⁵Department of Pathology, Medical University of Innsbruck, Innsbruck, Austria⁶Department of Medicine-IV, Medical University of Innsbruck, Innsbruck, Austria**ABSTRACT**

The majority of published cases of Megacystis Microcolon Intestinal Hypoperistalsis Syndrome (MMIHS), a disorder of smooth muscle function of hollow visceral organs is associated with mutations in the actin gamma2 (*ACTG2*) gene on chromosome 2p13. While alterations in smooth muscle actin histology have been described for the gastrointestinal tract no such information is available for urogenital organs of patients with *ACTG2* mutation-based MMIHS. We report the first description of actin gamma2 pathohistology in bladder smooth muscle of a male MMIHS-patient bearing a *de novo* *ACTG2*-mutation (c.770G>A; p.R257H) who suffered from voiding insufficiency due to severely deficient bladder contractility as demonstrated by video-urodynamic investigations. Our findings are a further contribution to the elucidation of pathogenic mechanisms of functional myopathic bladder atony.

Key Words: *ACTG2*-mutation, functional bladder atony, MMIHS, urology, myopathy**✉ Peter Heinz-Erian**

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Introduction

Megacystis Microcolon Intestinal Hypoperistalsis Syndrome (MMIHS) is a myopathic disorder caused by a variety of genetic defects affecting the smooth muscle cell contraction apparatus of both the urogenital and gastrointestinal tracts [1]. Clinically, MMIHS is characterized by megacystis (bladder distension in

the absence of mechanical obstruction), microcolon and intestinal hypoperistalsis [1,2]. Presently, the most thoroughly characterized underlying pathogenic mechanism of MMIHS is that of actin gamma2 dysfunction which has been associated with mutations in the actin gamma 2 (*ACTG2*) gene [3]. For the gastrointestinal tract, reports on patients with MMIHS provide pathohistological information on granular abnormalities in actin gamma 2 filament formation, subcellular localization and organization in smooth muscle cells as well as on functional consequences in terms of smooth muscle contraction [4,5].

While smooth muscle actin alterations have been published in the urinary tract of patients with MMIHS of unknown genetic origin [6], no smooth muscle histology of the bladder has been reported for *ACTG2* mutation-associated MMIHS to date. We describe for the first time

histopathological and immunohistochemical changes in smooth muscle actin gamma2 structure in the urinary tract of a patient suffering from MMIHS associated with a *de novo* heterozygous mutation (c.770G>A; p.R257H) in the *ACTG2* gene.

Case presentation

The male patient was born at term to a couple of Serbian caucasian origin who were fourth degree cousins. During his first years of life he suffered from predominantly gastrointestinal problems including intractable constipation which led to repeated partial resections of the ileum and colon eventually resulting in subtotal ileo-colectomy. Hirschsprung's disease was histologically excluded. During the following years the patient was frequently hospitalized because of repeated episodes of electrolyte and acid-base imbalance following intermittent bouts of severe diarrhea. After multiple complications including adhesion ileus causing further laparotomies, progressive intestinal failure and several urinary tract infections, the 17-year-old adolescent was admitted to our hospital for further diagnostic evaluation.

He presented with severe dehydration, cachexia (weight 36 kg, height 157 cm), bloated abdomen, watery diarrhea and hypochloremic alkalosis. Congenital Chloride Diarrhea (CCD) was suspected, however, mutational analysis of the *SLC26A3* gene was normal, excluding this disorder. During the subsequent six months the patient was successfully weaned from parenteral nutrition and was able to maintain near normal oral alimentation with frequent meals (eight to ten times daily) and substitution of large amounts of

oral fluid, NaCl (upto 32 mmol/kg/d) and KCl (upto 20 mmol/kg/d). Two years later the patient's weight was 54 kg, his height was 173 cm. Under this regimen he was able to maintain rather good health, to successfully conclude his college graduation exams and obtain his driver's license. He even managed to drive his car on his own for more than 1000 km from his hometown to our hospital for medical control visits.

From the age of 23 years on he suffered from increasingly frequent bouts of urinary tract infections. Increased bladder volume, and a completely atonic bladder with no increase in detrusor pressure were diagnosed by ultrasound and video-urodynamic investigations (Fig 1). Radiographic images showed atonic bladder contours with unsatisfactory spontaneous emptying even on deliberate exertion of manual abdominal pressure. Transurethral resection of the prostate and bladder neck was performed. Light microscopy of bladder neck tissue showed atrophic smooth muscle cells surrounded by reticular fibrosis shown in Figs 2a and 2b as well as decreased and irregular granular smooth muscle actin expression by immunohistochemistry can be viewed in Fig 2c. There were also signs of chronic nonspecific inflammation but no discernible pathology of the neural network. Thinning and fibrosis of the smooth muscle in the internal bladder sphincter were similar to those found in full thickness biopsies from the patient's rectum (not shown). Together with the findings of intestinal hypoperistalsis this suggested MMHIS, a hollow visceral myopathy, a diagnosis that was finally confirmed by the genetic finding of a *de novo* mutation in the *ACTG2* gene (c.770G>A; p.R257H) on chromosome 2p13.1.

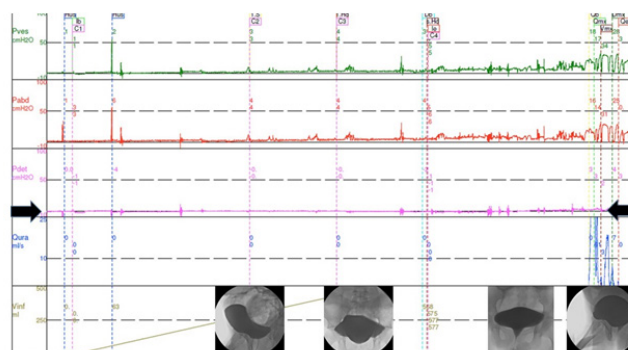


Fig. 1. Videourodynamic examination: Completely atonic bladder with no increase in detrusor pressure (black arrows) up to a filling of >1000 ml. Radiographic images showing atonic bladder contours with unsatisfactory spontaneous emptying utilizing abdominal pressure after liberal bladder neck incisions.

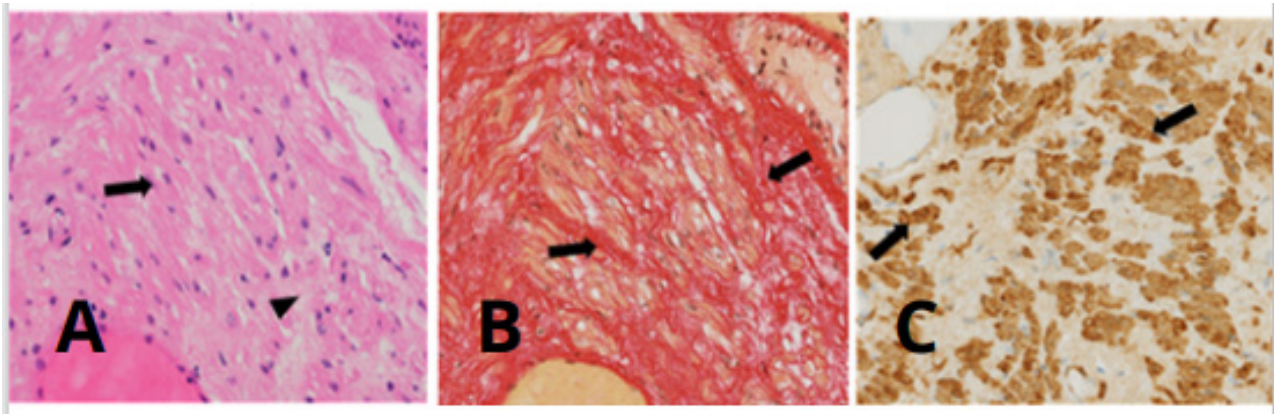


Fig. 2. Bladder neck tissue (x400), (A) H and E stain showing atrophy and intracellular vacuolation of smooth muscle cells (arrow) and increased intercellular fibrosis (arrowhead) (B) Picosirius Red stain indicating massive reticular fibrosis (arrows) (C) SMA II immunohistochemistry showing irregular granular smooth muscle actin expression.

Further frequent urinary tract infections occurred even after a perineostoma to facilitate emptying of the bladder had been applied. Renal function gradually deteriorated and renal dialysis had to be initiated soon after his 28th birthday. The possibility of multivisceral transplantation including kidneys and intestine was considered but rejected because of widespread colonization with multiresistant bacteria. The patient died shortly after his 30th birthday in the course of another severe bout of urosepsis in his hometown hospital.

Results and discussion

Megacystis and bladder atony are key features of MMIHS and are often associated with hydronephrosis, frequent urinary tract infections and renal insufficiency [7]. In our patient, apart from a few urinary tract infections which had been successfully treated without any problem on all occasions, no megacystis was diagnosed before the age of 22 years. Considering that the majority of MMIHS patients are diagnosed with megacystis and/or hydronephrosis prenatally or during the first year of life [8], our patient's disease course was very unusual and cannot be explained by the available information. Even at the age of 22 years an ultrasound scan of the bladder wall was reported to be unremarkable although hydronephrosis had been diagnosed before. Two years later, however, a voiding cysto-urethrogram demonstrated a large-capacity bladder with one liter of residual volume of urine which did not improve after bladder neck dilatation and not even after bladder neck

resection.

The latter procedure allowed us to obtain smooth muscle-containing tissue from the internal sphincter for microscopic evaluation which by demonstrating irregular smooth muscle actin expression shed further light on the probable mechanism of the patient's defective bladder function. Unfortunately, we were unable to obtain bladder wall tissue for more specifically defining actin gamma2 pathohistology in the detrusor muscle, which harbours the main effector mechanism for bladder evacuation. Nevertheless, our observation of actin gamma2 deficiency in the internal bladder sphincter, the structure that normally constricts the internal orifice of the urethra but is non-functional in MMIHS, may provide an important clue regarding the patient's voiding inefficiency. This latter mechanism may, besides stasis of urine in the bladder, also have been one factor in our patient's propensity for frequent urinary tract infections caused by bacteria ascending from the penile orifice.

Thus far, to the best of our knowledge, smooth muscle actin gamma2 pathohistology in the atonic bladder of a patient with MMIHS associated with a *de novo* heterozygous mutation in the *ACTG2* gene has not been reported and will thus provide further insight into the underlying mechanisms of myopathic bladder atony.

Conclusion

This course of our patient's disease, besides typical early onset gastrointestinal symptomatology, was characterized by uro-nephrologic dysfunction that

became manifest at an unusually late age. Our report on deficient actin pathohistology in the bladder provides additional information on the mechanism of disordered voiding in *ACTG2*-mutation-based MMIHS and is a further step in unraveling myopathic mechanisms of functional bladder atony.

Consent

Patient consent form has been obtained.

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